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Predation of Juvenile and Adult Anurans by Invertebrates: Current Knowledge and Perspectives

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Anuran amphibians are preyed on by vertebrates, invertebrates, and even carnivorous plants (Duellman and Trueb 1994). Most of these reports on predation are anecdotal (Fitch 1987; Greene 1993) and do not provide data other than a short description of the predatory event (e.g., Boistel and Pauwels 2002; Brandão and Garda 2000; Del-Grande and Moura 1997; Mitchell 1990). The scattered information on the subject makes it difficult to identify patterns. Additionally, McCormick and Polis (1982) pointed out the lack of quantitative data evaluating the impact of arthropod predators upon vertebrates. This is particularly true for predation by invertebrates upon post-metamorphic (generally adult) anurans. For example, reports on this subject usually state that few cases of invertebrate predation upon anurans are recorded (e.g., Bastos et al. 1994; Bernarde et al. 1999; Del-Grande and Moura 1997; Hinshaw and Sullivan 1990; Mitchell 1990), when, in fact, a considerable amount of information is generally available (e.g., McCormick and Polis 1982). Therefore, I review the subject in an attempt to depict our current knowledge, add unpublished data, and provide a background to which new reports may be added.

By reviewing published information on the subject I collected data on a wide range of taxa, i.e., at least 68 post-metamorphic (juvenile to adult) anuran species preyed upon by at least 57 invertebrate species, including arachnids, crabs, leeches, and various insect groups (Tables 1 and 2). Besides the species listed (Table 2), there exists indirect evidence and laboratory studies that add

TABLE 1. Invertebrate predators and number of species reported to prey upon post-metamorphic anuran amphibians.

Class	Order	Family	Common name	Abbreviation	Number Of species			
Hirudinea	Arhynchobdellida	Hirudinidae	Leeches	Ah	1			
Chilopoda	Scolopendromorpha	Scolopendridae	Giant centipedes	Gc	1			
Arachnida	Scorpiones	Buthidae	Scorpions	Sc	1			
		Uropygi	Vinegaroons	Ut	1			
	Araneae	Amblypygi	Amblypygidae	Amblypygids	Am	1		
		Araneidae	Araneidae	Orb weavers	Aa	3		
			Ctenidae	Wandering spiders	Ac	2		
			Ctenizidae	Trapdoor spiders	Az	1		
			Dipluridae	Tarantulas	Ad	1		
			Lycosidae	Wolf spiders	Al	6		
			Pisauridae	Fishing spiders	Ap	10		
			Sparassidae	Crab spiders	As	1		
			Theraphosidae	Tarantulas	At	6		
			Malacostraca	Decapoda	Coenobitidae	Crabs	Dc	1
			Hexapoda	Coleoptera	Carabidae	Ground beetles	Cb	3
					Cicindelidae	Tiger beetles	Cc	1
Dytiscidae	Diving beetles	Cd			1			
Diptera	Tabanidae	Horse flies			Di	1		
Hemiptera	Belostomatidae	Belostomatidae		Water bugs	Hb	8		
		Nepidae		Water scorpions	Hn	1		
	Hymenoptera	Formicidae		Ants	Hf	4		
Mantodea	Mantidae	Preying mantis		Mm	1			
Neuroptera	Corydalidae	Hellgrammites		Ch	1			

other potential invertebrate predators to the list, such as spiders (*Oilos antaguensis*, *Stasina portoricensis*, and *Avicularia latea*), amblypygids (*Phrynus longipes*), and forest crabs (*Epilobocera situatifrons*) (Formanowicz et al. 1981; Stewart 1995). Laboratory studies using pipid and hyperoliid frogs report pisaurid spiders as additional potential predators upon previously unreported anuran families (Table 2).

It appears that many anuran species can be preyed upon by invertebrate predators, independent of prey body size/age (see discussion in McCormick and Polis 1982), phylogeny, or recognized presence of an elevated quantity of biologically active skin secretions (Duellman and Trueb 1994) (Table 2). However, the risk of predation by invertebrates seems to be greater in two crucial periods of the anurans life cycle: 1) during the breeding season, when most species enter the water and consequently are in contact with potential aquatic predators (e.g., Bastos et al. 1994; Haddad and Bastos 1997; Toledo 2003); and 2) when the recently-metamorphosed frogs are about to leave or actually leave the water (Fig. 1), thus facing both aquatic and terrestrial invertebrates (e.g., Clerke and Williamson 1992; Hirai and Hidaka 2002; Robertson 1989; Toledo 2003). Almost 90% of the observations that provide descriptions of the frog behavior before the predation were recorded during these two stages.

Predation events occurred both in and out of the water and about 73% of them involved water bugs (approximately 25%) and spiders (approximately 48%) as anuran predators (Fig. 2). This may reflect the high density of these animals in nature (DuBois and Gobin 2001; Formanowicz et al. 1981; McCormick and Polis 1982; and references therein). Additionally, it could indicate that spiders and water bugs may be significant predators of anuran popu-

lations (Formanowicz et al. 1981; Toledo 2003), though, few studies have determined actual predation rates on adult anurans (e.g., Haddad and Bastos 1997; Hinshaw and Sullivan 1990).

In reviewing the subject I was able to identify a few cases of incorrect or repeated data. For example, Nauman and Dettlaff (1999) reported "the first published record of a giant water bug preying on an adult frog"; however, at least three reports on giant water bugs preying on adult frogs were already available by that time (Bastos et al. 1994; Haddad and Bastos 1997; Hinshaw and Sullivan 1990). Additionally, both Toledo (2003) and Brasileiro



FIG. 1. Juvenile *Hyla albosignata* being preyed upon by a tarantula on vegetation near a stream in a forested area, Municipality of Pilar do Sul, State of São Paulo, Brazil. Photograph by André Antunes.

TABLE 2. Post-metamorphic (juvenile to adult) anurans (15 families; at least 68 species) reported as prey of invertebrates (22 families; at least 57 species) and the microhabitat where the predation occurred. Anuran specific names follow Frost (2004), and thus some genera and species are updated. Predators' abbreviations are in table 1. An asterisk (*) after the anuran names indicates the recognized presence of a high amount of biologically active skin secretions.

Anurans (Prey)	Invertebrates (Predators)	Predation microhabitat	References
Ascaphidae			
<i>Ascaphus truei</i>	Hellgrammite – unidentified (Mh)	Pool in the stream	Jones and Raphael, 1998
Bufoiidae			
<i>Bufo bufo</i> *	<i>Formica rufa</i> (Hf)	Lake margin	Zuffi, 2001
<i>Bufo crucifer</i> *	<i>Lethocerus grandis</i> (Hb)	Temporary pond	Haddad and Bastos, 1997
<i>Bufo houstonensis</i> *	<i>Solenopsis invicta</i> (Hf)	Litter	Thomas and Allen, 1997
<i>Bufo marinus</i> *	<i>Iridomyrmex purpureus</i> (Hf)	Pond margin	Clerke and Williamson, 1992
<i>Bufo marinus</i> *	<i>Scolopendra alternans</i> (Gc)	Leaf litter	Carpenter and Gillingham, 1984
<i>Bufo terrestris</i> *	<i>Lethocerus</i> sp. (Hb)	Temporary pond	McCoy, 2003
Centrolenidae			
<i>Centrolene prosoblepon</i>	<i>Cupiennus</i> sp. (Ac)	Over rocks, near the water	Hayes, 1983
<i>Hyalinobatrachium fleischmanni</i>	<i>Cupiennus</i> sp. (Ac)	Over leaf	Hayes, 1983
Dendrobatidae			
<i>Colostethus inguinalis</i>	Freshwater crab – unidentified (Dc)	Not provided	Duellman and Trueb, 1994
<i>Dendrobates auratus</i> *	<i>Sericopelma rubronitens</i> (At)	Litter	Summers, 1999
<i>Dendrobates pumilio</i> *	<i>Paraponera clavata</i> (Hf)	Litter	Fritz et al., 1981
Hylidae			
<i>Acris crepitans</i>	<i>Hogna helluo</i> (Al)	Semi-permanent wetland	Blackburn et al., 2002
<i>Acris gryllus</i>	<i>Dolomedes</i> sp. (Ap)	Edges of water body	Goin, 1943
<i>Hyla albomarginata</i>	<i>Belostoma</i> sp. (Hb)	Temporary pond	Froehlich, 2001
<i>Hyla albosignata</i>	Tarantula – unidentified (At)	Vegetation over water	A. Antunes, unpubl. data
<i>Hyla cinerea</i>	<i>Dolomedes okefinokensis</i> (Ap)	Vegetation over water	Jeffery et al., 2004
<i>Hyla cinerea</i>	<i>Acanthepeira stellata</i> (Aa)	Not provided	Lockley, 1990
<i>Hyla crepitans</i>	<i>Belostoma</i> sp. (Hb)	Permanent pool	Mijares-Urrita et al., 1997
<i>Hyla ebraccata</i>	<i>Cupiennius coccineus</i> (Ac)	Swamp	Szelistowski, 1985
<i>Hyla japonica</i>	<i>Diplonychus japonicus</i> (Hb)	Flooded rice field	T. Hirai, unpubl. data
<i>Hyla japonica</i>	<i>Dolomedes sulfurous</i> (Ap)	Flooded rice field	T. Hirai, unpubl. data
<i>Hyla japonica</i>	<i>Laccotrephes japonensis</i> (Hn)	Flooded rice field	T. Hirai, unpubl. data
<i>Hyla japonica</i>	<i>Lethocerus deyrollei</i> (Hb)	Flooded rice field	Hirai and Hidaka, 2002
<i>Hyla jimi</i>	<i>Belostoma elongatum</i> (Hb)	Temporary pond	Toledo, 2003
<i>Hyla minuta</i>	<i>Belostoma elongatum</i> (Hb)	Temporary pond	Toledo, 2003
<i>Hyla minuta</i>	<i>Lethocerus delpontei</i> (Hb)	Permanent pond	Bastos et al., 1994
<i>Hyla minuta</i>	<i>Dolomedes</i> sp. (Ap)	Permanent pond	Bastos et al., 1994
<i>Hyla minuta</i>	<i>Ancylometes vulpes</i> (Ap)	Permanent pond	Bernarde et al., 1999
<i>Hyla minuta</i>	<i>Ancylometes gigas</i> (Ap)	Permanent pond	Bernarde et al., 1999
<i>Hyla miotypanum</i>	<i>Abedus</i> sp. (Hb)	Permanent stream	Pineda, 2003
<i>Hyla nana</i>	<i>Thaumasia</i> sp. (Ap)	Swamp	Pramuk and Alamillo, 2002
<i>Hyla sanborni</i>	<i>Diapontia cf. uruguayensis</i> (Al)	Web over pond	Del-Grande and Moura, 1997
<i>Hyla versicolor</i>	<i>Argiope aurantia</i> (Aa)	Web near pond	Steehouder, 1992
<i>Hyla versicolor</i>	<i>Lethocerus americanus</i> (Hb)	Pond	Hinshaw and Sullivan, 1990
<i>Litoria caerulea</i> ~	<i>Hierodula wernerii</i> (Mm)	Not provided	Ridpath, 1977
<i>Litoria caerulea</i>	<i>Atrax formidabilis</i> (Ad)	Not provided	McCormick and Polis, 1982
<i>Litoria ewingi</i>	<i>Catadromus lacordairei</i> (Cb)	Not provided	LittleJohn and Wainer, 1978
<i>Litoria lesueurii</i>	<i>Lycosa lapidosa</i> (Al)	On the rocks of a creek bed	Raven, 1990
<i>Litoria raniformis</i>	<i>Archimantis latistyla</i> (Mm)	Not provided	Ridpath, 1977
<i>Pseudacris crucifer</i>	Diving beetle – unidentified (Cd)	Temporary pond	Hinshaw and Sullivan, 1990
<i>Pseudacris feriarum</i>	<i>Dolomedes triton</i> (Ap)	Temporary pond	Mitchell, 1990
<i>Pseudacris ocularis</i>	<i>Lycosa</i> sp. (Al)	Ground, near water body	Owen and Johnson, 1997
<i>Scinax alter</i>	<i>Ancylometes rufus</i> (Ap)	Over aquatic vegetation	Prado and Borgo, 2003
<i>Scinax alter</i>	<i>Thaumasia</i> sp. (Ap)	Water surface	Marra et al., 2003
<i>Scinax cruentommus</i>	Wolf spider – unidentified (Al)	Vegetation over ground	Aucone and Card, 2002
<i>Scinax elaeochroa</i>	<i>Cupiennius coccineus</i> (Ac)	Swamp	Szelistowski, 1985
<i>Scinax fuscomarginatus</i>	<i>Oxyurptychus brasiliensis</i> (Ah)	Vegetation over pond	Brand, o and Garda, 2000
<i>Scinax fuscomarginatus</i>	Tarantula – unidentified (At)	Temporary pond	L. F. Toledo, unpubl. data
<i>Scinax fuscomarginatus</i>	Water bug – unidentified (Hb)	Temporary pond	L. F. Toledo, unpubl. data
<i>Scinax ruber</i>	Preying mantis – unidentified (Mm)	Over vegetation	J. L. Guillaumet, unpubl. data
<i>Scinax squairostris</i>	<i>Belostoma elongatum</i> (Hb)	Temporary pond	Toledo, 2003

TABLE 2. Continued.

Anurans (Prey)	Invertebrates (Predators)	Predation microhabitat	References
<i>Scinax</i> sp. (<i>aff. similis</i>)	<i>Belostoma elongatum</i> (Hb)	Temporary pond	Toledo, 2003
Hyperoliidae			
<i>Hyperolius marmoratus</i>	<i>Thalassius fimbriatus</i> (Ap)	Laboratory situation	McCormick and Polis, 1982
Leptodactylidae			
<i>Eleutherodactylus coqui</i>	<i>Olios</i> sp. (As)	Above ground	Formanowicz Jr. et al., 1981
<i>Eleutherodactylus coqui</i>	<i>Oligothenus ottleyi</i> (Al)	Not provided	Formanowicz Jr. et al., 1981
<i>Eleutherodactylus coqui</i>	<i>Phrynos palmatus</i> (Am)	Not provided	Formanowicz Jr. et al., 1981
<i>Eleutherodactylus coqui</i>	<i>Tityus obtusus</i> (Sc)	Vegetation over ground	Villanueva-Rivera et al., 2000
<i>Eleutherodactylus zugi</i>	<i>Ctenus vernalis</i> (Ac)	Ground inside cave	Novo et al., 1985
<i>Eleutherodactylus</i> sp.	<i>Paraponera clavata</i> (Hf)	Litter	Fritz et al., 1981
<i>Eleutherodactylus</i> spp.	<i>Cupiennius coccineus</i> (Ac)	Experimental condition	Szelistowski, 1985
<i>Hylodes phyllodes</i>	<i>Trachalea keyserlingi</i> (Ap)	Leaf litter next to a stream	Schiesari et al., 1995
<i>Leptodactylus knudseni</i> *	<i>Theraphosa leboni</i> (At)	Not provided	Boistel and Pauwels, 2002
<i>Leptodactylus labyrinthicus</i> *	<i>Belostoma elongatum</i> (Hb)	Temporary pond	Toledo, 2003
<i>Leptodactylus ocellatus</i>	<i>Lethocerus annulipes</i> (Hb)	Not provided	Lima, 1940
<i>Physalaemus cuvieri</i>	<i>Belostoma elongatum</i> (Hb)	Temporary pond	Toledo, 2003, Brasileiro et al., 2003
<i>Physalaemus fuscomaculatus</i> *	<i>Belostoma elongatum</i> (Hb)	Temporary pond	Toledo, 2003
<i>Physalaemus cf. fuscomaculatus</i> *	<i>Lethocerus</i> sp. (Hb)	Temporary pond	Giaretta and Menin, 2004
<i>Physalaemus pustulosus</i>	<i>Sericopelma rubronitens</i> (At)	Leaf litter	Gray et al., 1999
<i>Physalaemus spiniger</i>	Wolf spider – unidentified (Al)	Temporary pond	L. M. Giasson, unpubl. data
Limnodynastidae			
<i>Limnodynastes tasmaniensis</i>	<i>Catadromus lacordairei</i> (Cb)	Not provided	LittleJohn and Wainer, 1978
<i>Neobatrachus centralis</i>	<i>Selenotypus</i> sp. (At)	Not provided	Raven, 1990
<i>Neobatrachus centralis</i>	<i>Selenocosmia crassipes</i> (At)	Not provided	McCormick and Polis, 1982
Microhylidae			
<i>Microhyla ornata</i>	<i>Lycosa carmichaeli</i> (Al)	Not provided	McCormick and Polis, 1982
Myobatrachidae			
<i>Crinia pseudinsignifera</i>	<i>Aganippe raphiduca</i> (Az)	Ground, near spider burrow	Butler and Main, 1959
<i>Crinia signifera</i>	<i>Chlaenius darlingensis</i> (Cb)	Margin of pond	Robertson, 1989
<i>Uperoleia laevigata</i>	<i>Chlaenius darlingensis</i> (Cb)	Margin of pond	Robertson, 1989
Pelobatidae			
<i>Spea multiplicata</i>	<i>Tabanus punctifer</i> (Di)	Mud margin of pond	Jackman et al., 1983
<i>Spea multiplicata</i>	<i>Cicindela sedecimpunctata</i> (Cc)	Not provided	McCormick and Polis, 1982
Pipidae			
<i>Xenopus laevis</i> *	<i>Dolomedes triton</i> (Ap)	Laboratory situation	Rogers, 1996
Rhacophoridae			
<i>Rhacophorus arboreus</i>	<i>Cybister japonicus</i> (Cd)	Flooded rice field	T. Hirai, unpubl. data
<i>Rhacophorus schlegelii</i>	<i>Laccotrephes japonensis</i> (Hn)	Flooded rice field	T. Hirai, unpubl. data
<i>Rhacophorus schlegelii</i>	<i>Lethocerus deyrollei</i> (Hb)	Flooded rice field	Hirai and Hidaka, 2002
Ranidae			
<i>Euphylyctis cf. cyanophlyctis</i>	<i>Lycosa barmanica</i> (Al)	Not provided	McCormick and Polis, 1982
<i>Fejervarya limnocharis</i>	<i>Lethocerus deyrollei</i> (Hb)	Flooded rice field	Hirai and Hidaka, 2002
<i>Rana cascadae</i>	<i>Lethocerus</i> sp. (Hb)	Lake	Nauman and Dettlaff, 1999
<i>Rana clamitans</i>	Wolf Spider – unidentified (Al)	Grass field	Neil, 1948
<i>Rana nigromaculata</i>	<i>Epomis nigricans</i> (Cb)	Flooded rice field	T. Hirai, unpubl. data
<i>Rana nigromaculata</i>	<i>Lethocerus deyrollei</i> (Hb)	Flooded rice field	Hirai and Hidaka, 2002
<i>Rana porosa</i>	<i>Lethocerus deyrollei</i> (Hb)	Irrigation ditch (rice field)	T. Hirai, unpubl. data
<i>Rana rugosa</i>	<i>Lethocerus deyrollei</i> (Hb)	Flooded rice field	T. Hirai, unpubl. data
Unidentified anurans			
Frog	<i>Grammostola</i> sp. (At)	Not provided	McCormick and Polis, 1982
Frog	<i>Lasidora</i> sp. (At)	Not provided	McCormick and Polis, 1982
Frog	<i>Birgus latro</i> (Dc)	Not provided	McCormick and Polis, 1982
Frogs and toads	<i>Mastigoproctus giganteus</i> (Ut)	Not provided	McCormick and Polis, 1982
Green Frog	<i>Nephila plumipes</i> (Aa)	Not provided	McCormick and Polis, 1982
Green Tree Frog	<i>Nephila plumipes</i> (Aa)	Not provided	McCormick and Polis, 1982
Leptodactylidae, Brown Frogs	Wolf Spider – unidentified (Al)	Not provided	McCormick and Polis, 1982
Tree Frog	<i>Dolomedes okefenokensis</i> (Ap)	Not provided	McCormick and Polis, 1982

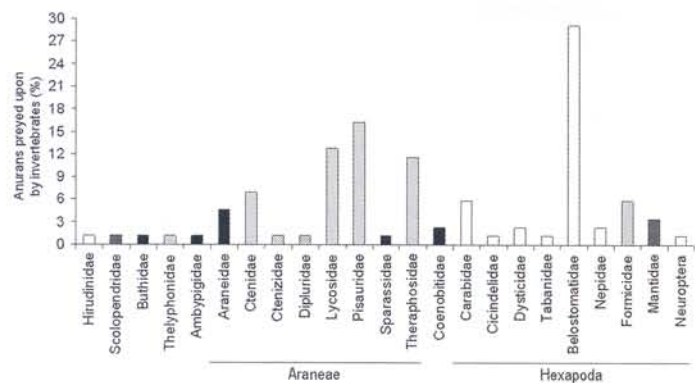


FIG. 2. Percentage of different anuran species that were preyed upon by different invertebrate families based on references listed in Table 2 ($N_{\text{total}} = 89$ accounts). White bars indicate predation events reported to occur in the water; dark bars indicate predation events out of the water; gray bars indicate predation events both in and out of the water; and striped bars represent lack of information on the microhabitat in which the predation occurred.

et al. (2003) provided duplicate reports of the predation of *Physalaemus cuvieri* by the same water bug species (*Belostoma elongatum*) at the same study site. Nevertheless, repeated records may be beneficial as they provide more evidence for an actual predator-prey relationship, and may help determine if any geographic variation occurs in the predator-prey relationship.

Despite the considerable number of reports much more information is likely to appear in the next few years. However, the simple descriptions of a predatory event without providing further details (e.g., microhabitat and prey activity before predation - data that should generally be available to the observers) make future discussions and predictions difficult (see Greene 1993). Therefore, even reports on simple interactions between predator and prey (particularly in the context discussed here) should provide more detailed accounts whenever possible (see complementary discussions in Greene 1986; 1993).

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**Application and Evaluation of a Stomach Flushing
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Crocodylian diets can be studied by observing what an animal eats, conducting feeding trials on captive animals, performing biochemical and isotope analysis, or by obtaining samples of the ingested food from the stomachs of wild animals. Stomach contents can be obtained post-mortem from specimens killed for that purpose or collected incidentally from commercial harvests. However, many crocodylian species are threatened or endangered and there are ethical and practical constraints on killing animals for study. Therefore, non-lethal methods have been developed to obtain stomach contents from live crocodylians without causing them harm.

Non-lethal methods used to obtain the stomach contents of crocodylians fall into three categories: invasive scoops that mechanically retrieve material through the esophagus (Taylor et al. 1978), irrigation methods that introduce water and flush material from the stomach (Fitzgerald 1989; Taylor et al. 1978) and combinations of these (Webb et al. 1982). These methods vary in effectiveness at recovering all the stomach contents, ease of application in the field and degree of invasive trauma to the animal. The purpose of this study is to evaluate the hose-Heimlich technique (irrigation method) for accuracy and to check for internal damage on American Alligators (*Alligator mississippiensis*). We also report on the effectiveness of the hose-Heimlich technique used during an alligator diet study, and we investigate alternative water sources for use with this technique.

The hose-Heimlich technique described by Fitzgerald (1989) was used to obtain the stomach contents from live adult American

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